**INDOLE TEST REAGENTS**

<table>
<thead>
<tr>
<th>Cat. no.</th>
<th>Description</th>
<th>Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>Z65</td>
<td>Indole Spot Reagent</td>
<td>15ml</td>
</tr>
<tr>
<td>Z67</td>
<td>Indole Kovacs Reagent</td>
<td>15ml</td>
</tr>
</tbody>
</table>

**INTENDED USE**

Hardy Diagnostics Indole Spot Reagent and Indole Kovacs Reagent are recommended for use in determining the indole reaction of bacteria.

**SUMMARY**

The indole test is a qualitative procedure for determining the ability of bacteria to produce indole by deamination of tryptophan.

Using Kovacs tube method, indole combines, in the presence of a tryptophan rich medium, with p-Dimethylaminobenzaldehyde at an acid pH in alcohol to produce a red-violet compound.

In the spot test, indole combines, in the filter paper matrix, at an acid pH with p-Dimethylaminocinnamaldehyde (DMACA) to produce a blue to blue-green compound. Indole Spot Reagent (DMACA) has been reported to be useful in detecting indole production by members of the family *Enterobacteriaceae* and certain anaerobic species.

**REAGENT FORMULA**

Ingredients per liter:*

![Indole Spot Reagent](https://catalog.hardydiagnostics.com/cp_prod/Content/hugo/IndoleTestRgnts.htm)

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>p-Dimethylaminocinnamaldehyde (DMACA)</td>
<td>10.0gm</td>
</tr>
<tr>
<td>Hydrochloric Acid, 37%</td>
<td>100.0ml</td>
</tr>
<tr>
<td>Deionized Water</td>
<td>900.0ml</td>
</tr>
</tbody>
</table>

**Indole Kovacs Reagent:**

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>p-Dimethylaminobenzaldehyde</td>
<td>50.0gm</td>
</tr>
<tr>
<td>Hydrochloric Acid, 37%</td>
<td>250.0ml</td>
</tr>
<tr>
<td>Amyl Alcohol</td>
<td>750.0ml</td>
</tr>
</tbody>
</table>

* Adjusted and/or supplemented as required to meet performance criteria.

**STORAGE AND SHELF LIFE**
Storage: Upon receipt store at 2-30°C. Products should not be used if there are any signs of deterioration or if the expiration date has passed.

The expiration dating on the product label applies to the product in its intact packaging when stored as directed.

Refer to the document "Storage" on the Hardy Diagnostics Technical Document website for more information.

**PRECAUTIONS**

This product may contain components of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not guarantee the absence of transmissible pathogenic agents. Therefore, it is recommended that these products be treated as potentially infectious, and handle observing the usual universal blood precautions. Do not ingest, inhale, or allow to come into contact with skin.

This product is for *in vitro* diagnostic use only. It is to be used only by adequately trained and qualified laboratory personnel. Observe approved biohazard precautions and aseptic techniques. All laboratory specimens should be considered infectious and handled according to "standard precautions." The "Guidelines for Isolation Precautions" is available from the Centers for Disease Control and Prevention at [www.cdc.gov/ncidod/dhqp/gl_isolation.html](http://www.cdc.gov/ncidod/dhqp/gl_isolation.html).

For additional information regarding specific precautions for the prevention of the transmission of all infectious agents from laboratory instruments and materials, and for recommendations for the management of exposure to infectious disease, refer to CLSI document M-29: *Protection of Laboratory Workers from Occupationally Acquired Infections: Approved Guideline*.

Sterilize all biohazard waste before disposal.

Refer to the document "Precautions When Using Media" on the Hardy Diagnostics Technical Document website for more information.

Refer to the document SDS Search instructions on the Hardy Diagnostics website for more information.

**PROCEDURE**

Specimen Collection: This product is not intended for primary isolation of patient specimens. This product is used in conjunction with other biochemical tests to identify cultures of isolated organisms.

**Indole Spot Reagent (DMACA):** Place several drops of Indole Spot Reagent on a piece of filter paper. With an inoculating loop or wooden applicator stick, pick a portion of an 18-24 hour isolated colony from a non-selective media and rub it onto the reagent saturated area of the filter paper.

Interpretation of Results: A positive reaction is denoted by the appearance of a blue to blue-green color change on the bacterial smear, or red-violet in the case of *Providencia alcalifaciens*, within 10 seconds. Negative reactions remain colorless or light pink.

The filter paper may appear pink to purple after the reagent is applied, but only the color change of the bacterial smear itself should be indicative of a positive reaction.

**Indole Kovacs Reagent:** Lightly inoculate Tryptone Broth (Cat. no. R40) or Peptone Broth (Cat. no. K151) with the test organism. Incubate 24-48 hours at 35°C. Add 4-5 drops of Kovacs Reagent to the tube, shake gently.

Interpretation of Results: A positive Kovacs tube test reaction is denoted by the appearance of a pink to red color in the top alcohol layer. Negative reactions remain colorless or light yellow.

If Kovacs Indole is to be used with a commercial identification test strip, such as API ® or EnteroPluri, or Microgen, consult the manufacturer's literature.

**LIMITATIONS**

Indole tests may be used as an aid in the identification and differentiation of gram-positive and gram-negative organisms. Additional biochemical testing using pure cultures is recommended for complete identification.
The tube test is a more sensitive method of detecting indole than the spot test.

When performing a spot test, Kovacs Indole Reagent may be used as a substitute for the spot test reagent. However, Kovacs Indole Reagent, when used as the spot test reagent, is less sensitive in detecting indole than the Indole Spot Reagent (DMACA). (6)

Kovacs Indole Reagent is not recommended for use with anaerobic bacteria. The Indole Spot Reagent (DMACA) is suitable for anaerobe use.

Since peptones have been shown to vary with regard to their suitability for use with indole testing, media selected for indole determination should be tested with known positive and negative organisms to insure suitability.

Media containing glucose should not be used for indole testing due to the formation of acid end products which have been shown to reduce indole production. Mueller Hinton Agar should also not be used for this test because tryptophan is destroyed during acid hydrolysis of casein.

Media containing dye, such as MacConkey and EMB, are unsuitable sources of inoculum due to possible carryover of dye and subsequent interference of indole color interpretation.

Indole-positive colonies have been reported to cause adjacent indole-negative colonies to appear false-positive due to diffusion of indole into the media. To avoid false-positives, select colonies of different morphologies that are separated by at least 5mm for indole testing. (6)

**MATERIALS REQUIRED BUT NOT PROVIDED**

Standard microbiological supplies and equipment such as loops, Tryptone Broth (Cat. no. R40), Peptone Broth (Cat. no. K151), needles, incubators, and incinerators, etc., as well as biochemical and serological reagents, are not provided.

**QUALITY CONTROL**

Hardy Diagnostics tests each lot of commercially manufactured media using appropriate quality control microorganisms and quality specifications as outlined on the Certificates of Analysis (CoA). The following organisms are routinely used for testing at Hardy Diagnostics:

<table>
<thead>
<tr>
<th>Test Organisms</th>
<th>Reaction</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Indole Spot</strong></td>
<td></td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>Positive; blue to blue-green color change</td>
</tr>
<tr>
<td>ATCC ® 25922</td>
<td></td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>Negative; no color change or pink</td>
</tr>
<tr>
<td>ATCC ® 27853</td>
<td></td>
</tr>
<tr>
<td><strong>Indole Kovacs</strong></td>
<td></td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>Positive; red color change</td>
</tr>
<tr>
<td>ATCC ® 25922</td>
<td></td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>Negative; no color change or light yellow</td>
</tr>
<tr>
<td>ATCC ® 27853</td>
<td></td>
</tr>
</tbody>
</table>

**USER QUALITY CONTROL**

End users of commercially prepared culture media should perform QC testing in accordance with applicable government regulatory agencies, and in compliance with accreditation requirements. Hardy Diagnostics recommends end users check for signs of contamination and deterioration and, if dictated by laboratory quality control procedures or regulation, perform quality control testing to demonstrate growth or a positive reaction and to demonstrate inhibition or a negative reaction, if applicable. Hardy Diagnostics quality control testing is documented on the certificates of analysis (CoA) available from Hardy Diagnostics Certificates of Analysis website. In addition, refer to
Showing positive indole reaction.
Filter paper was saturated with Indole Spot Reagent (Cat. no. Z65) and *Escherichia coli* (ATCC® 25922) growth was applied subsequently. *E. coli* was incubated aerobically for 24 hours at 35°C on a TSA plate (Cat. no. G60).

Showing negative indole reaction.
Filter paper was saturated with Indole Spot Reagent (Cat. no. Z65) and *Pseudomonas aeruginosa* (ATCC® 27853) growth was applied subsequently. *P. aeruginosa* was incubated aerobically for 24 hours at 35°C on a TSA plate (Cat. no. G60).

Showing positive indole reaction.
*Escherichia coli* (ATCC® 25922) was incubated in Tryptone Broth under aerobic conditions for 24 hours at 35°C. 5 drops of Indole Kovac's Reagent (Cat. no. Z67) was added directly to the broth and the tube was gently shaken. The top alcohol layer shows a positive reaction.

Showing negative indole reaction.
*Pseudomonas aeruginosa* (ATCC® 27853) was incubated in Tryptone Broth (Cat. no. R40) under aerobic conditions for 24 hours at 35°C. 5 drops of Indole Kovac's Reagent (Cat. no. Z67) was added directly to the broth and the tube was gently shaken. The top alcohol layer shows a negative reaction.

**REFERENCES**


   www.cms.hhs.gov/cla.

API is a registered trademark of bioMeriux, France.
ATCC is a registered trademark of the American Type Culture Collection.

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